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Full Length Article

Mifepristone Synchronization of Estrus and Timed Insemination for Swamp and River Crossbred Buffalo Heifers

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Abstract

The objectives of this study were to optimize the best dose of mifepristone injections in combination with PmPG synchronization protocols and the use of fixed–time artificial insemination (FTAI) to enhance the reproductive efficiency and the use of superior genetics of Chinese crossbred buffalo heifers. A total of 155 crossbred Buffalo heifers were used in this experiment, first 76 heifers were synchronized with PmPGMH in different doses (0.3, 0.4 and 0.5 mg/kg) of mifepristone. PmPGMH in mifepristone dose of 0.3 mg/kg showed the highest significant (P < 0.05) estrus rate (84.2%), ovulation rate (89.5%) and conception rate (52.6%) among other methods. Furthermore, 79 crossbred buffaloes were subjected to single FTAI using PmPGMH at h 16, 20, 24, 28 and 32 of mifepristone dose (0.3 mg/kg) treatment to select the best AI time, the results showed that TAI at h 24 was the best insemination time. © 2019 Friends Science Publishers

Key words: PMSG; Mifepristone; TAI; Buffalo heifers

Introduction

Buffalo is the most common domestic farm animal which can tolerate high temperature and humidity of tropical and subtropical countries so have been used for long time in Asia (Lu et al., 2015). It plays an important role in livestock production and the economy in china. However, the productivity of such animal is significantly diminished by poor reproductive efficiency represented in late puberty, seasonal breeding, low expression of estrus signs and difficult detection of ovulation time, higher fetal and embryonic mortality (Singh et al., 2000; Baruselli et al., 2001; Giuseppe et al., 2005). Silent estrus was one of the biggest problems in buffaloes which increased calving interval due to the failure of the estrous detection (Rajesh et al., 2017). That may be explained due to lower concentrations of hormones in buffalo than in cattle (Lokeshwar, 1990; Jainudeen and Hafez, 2016). Estrus detection greatly influenced the efficiency of artificial insemination programs in buffalo (Akhtar et al., 2014), so different protocols have been developed to synchronize follicular waves and ovulation in predetermined time to support FTAI, these protocols depend mainly on the combination of P4, E2, eCG (Rensis and López-Gatius, 2007; Carvalho et al., 2013, 2016, 2017; Monteiro et al., 2018), PMSG, GnRH and PGF2 (Turk et al., 2008;

Kumar et al., 2010).

PMSG treatment showed great efficiency in estrous synchronization and improved incidence of multiple ovulation in sheep and goat during the breeding season (Greyling and Niekerk, 1991; Mehaisen *et al.*, 2005; Ustuner *et al.*, 2007; Garoussi *et al.*, 2012). Moreover, it enhanced dairy cow ovarian follicular growth and fertility (Souza *et al.*, 2009; Rostami *et al.*, 2011). PMSG combination with CIDR resulted in estrus rate of 59.1% and conception rate of 66.6% and consequently improved fertility in true anoestrus buffaloes. However, PRID+PMSG treatment showed better ovarian follicular growth with average pregnancy rates of 70% (Presicce *et al.*, 2005; Kumar *et al.*, 2010; Caesar *et al.*, 2011).

Wide variation in estrus duration and low estrus expression resulted in difficult prediction of the time of ovulation so the use of AI for genetic improvement in buffaloes is limited (Baruselli, 1994; Baruselli, 2001). The use of these hormonal protocols can control follicular development and ovulation at a predetermined time to induce estrus in large percentage of females to allow TAI without estrus detection (Carvalho *et al.*, 2016; Monteiro *et al.*, 2018). This enable the beneficial use of AI for genetic improvement in buffalo industry (Berber, 2002).

Many recent studies has been performed on synchronization programs in buffaloes, however the average

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herd fertility remained low when compared to dairy cows (Arshad *et al.*, 2017). Moreover, FTAI estrus synchronization protocols gave un satisfactory results in heifers (Dahlen *et al.*, 2003; Lamb *et al.*, 2006). Lower pregnancy rates resulted from FTAI in heifers are mainly due to an inability to synchronize follicular waves with the same success that occurred in cows (Lamb *et al.*, 2006). Improving the reproductive performance of buffalo heifers became an important necessity in order to supply herd with suitable replacements, so attention needs to be focused on better understanding of the mechanisms controlling ovarian follicular growth and development for improving super ovulation responses and conception rates, and reducing embryonic mortality.

In order to optimize existing synchronization protocols in buffalo heifers, it is mandatory to improve the degree of induced synchronization between ovulation of dominant follicle and the time of AI, and to understand the fate of dominant follicle after the end of the synchronization protocol. Many synchronization protocols used a combination of exogenous hormones to regulate the estrous cycle (Muth-Spurlock et al., 2016), However, there is no information about the efficiency of mifepristone injection to promote ovulation in protocols of ovulation synchronization and TAI in buffalo. Mifepristone is a progesterone receptor (PR) antagonist, which works as antiprogestogen by blocking the effects of progesterone to induce ovulation (Carvalho et al., 2014). Thus, the current study aimed to evaluate the efficiency of combining mifepristone in different doses with PmPG synchronization protocols and to undertake a comprehensive evaluation of the response of Chinese crossbred buffalo heifers to this new estrus synchronization protocol. We hypothesized that PmPG treatment with mifepristone will induce a higher ovulation rate than PmPG synchronization protocols, thus increase herd fertility and reduce embryonic losses.

Materials and Methods

Experimental Site and Animal Care

All experimental protocols were approved by the Ethical Committee of the Hubei Research Center, Huazhong Agricultural University, China (Approval ID: SCXK (Hubei) 20080005).

Experimental Animals

A total of 155 crossbred buffalos (Mediterranean \times Murrah \times Nili Ravi \times Jianghan) were raised in a Buffalo Farm in Jinniu, Hubei province, China. The average ambient temperature varied from5 to 28 buffalo Farm in humidity ranging from 55% and 73% in breeding season (September to March) of year 2018. The body condition scores (BCS) of

buffaloes varied from points 2.5 to 3.5 at sexually mature stages, with mean body weight, chest girth, body height, body lengths and abdominal girths averaged 433.0 ± 72.1 kg, 186.9 ± 12.9 cm, 136.1 ± 5.8 cm, 128.9 ± 5.8 cm and 120.6 ± 5.5 cm, respectively. All the animals were healthy in regular estrus cycle and physical condition, fed with total mixed ration (TMR) consisted of forage (corn silage, rice straw, peanut vine) and concentrate (corn, wheat bran, soybean meal), and were machine—milked twice a day.

Experimental Groups

There were 2 trials in this study. Trial 1 was performed to optimize the dose of mifepristone in combination of PmPG with hCG in buffalo heifers. A total of 76 buffalo heifers were divided into 4 groups and were synchronized by PmPGMH protocol in 4 doses (0, 0.3, 0.4 and 0.5 mg/kg body weight) of mifepristone respectively. Mifepristone was injected at same time as the injection of GnRH (day 4). Artificial insemination was performed at the afternoon of day 5, which is 24 h after mifepristone injection. An injection of hCG (2000 IU) was given on day 5 after insemination.

Trial 2 was performed to optimize the time of AI in PmPGMH method. A total of 79 crossbred buffalo heifers were treated by PmPGMH protocols in 0.3mg/kg dose of mifepristone. Animals weredivided into 5 groups and given a single AI at h 16 (group 1), 20 (group 2), 24 (group 3), 28 (group 4), and 32 (group 5), respectively after mifepristone injection. An injection of hCG (2000 IU) was given on day 5 after insemination (Fig. 1).

Evaluation of Follicle Development and Ovulation

Follicle development was evaluated by ultrasonography on day $1^{\rm st}$ (1 day before PGF_{2 α} treatment) to $7^{\rm th}$ day (72 h after the injection of GnRH) by use of desktop B–type veterinary ultrasound scanner (WED–9618–v, equipped with LV2–3/6.5 MHz rectal probe, Shenzhen Well. D Medical Electronics Co., Ltd., Guangdong, China). Ultrasonography was performed twice a day at 6 am and 6 pm. Follicle dynamics were evaluated on the basis of follicles diameters. The time of sudden disappearance of dominant follicles were considered as ovulation to appear (Gimenes *et al.*, 2011; Liu *et al.*, 2016).

Estrus Detection and Pregnancy Diagnosis

Estrus was observed twice a day (6 a.m. and 6 p.m.) including the presence of vaginal mucus discharge, animal being mounted by others and the presence of > 9 mm follicle. Pregnancy diagnosis was performed by transrectal ultrasonography (Desktop B—type veterinary ultrasound scanner) day 40 after AI (Yindee *et al.*, 2011; Haider *et al.*, 2015; Liu *et al.*, 2016).

Statistical Analysis

Data were analyzed using statistical software package (S.P.S.S. version 17.0 for Windows). The ovulation rate, the growth speed of dominant follicles and the diameter of the ovulatory follicle were expressed as mean \pm standard error (SEM). Data for estrous rate, ovulation rate, follicular cyst rate and conception rate were calculated by the chi–squared test using Graph Pad Prism–6 software package (GraphPad Software Inc.), where P < 0.05 was considered as statistically significant. Data about follicle diameters were transformed firstly in logarithm (Fig. 3).

Results

Optimal dose of Mifepristone in PmPGMH Methods

Table 1 results showed that buffalo heifers treated by PmPGMH protocol in low dose (0.3 mg/kg) of mifepristone showed shorter (P > 0.05) estrus duration time, days of ovulation and initial follicle diameter (Table 1) and the biggest maximum follicle diameters than other groups (Fig. 2).

Table 2 results showed that higher estrous rate (84.2%), ovulation rate (89.5%), conception rate (47.4%), and the lowest silent estrus rate (0.5%) and follicular cysts rate (0.5%), although the follicle diameters at maximal size were not significantly different among all groups (Fig. 2). In comparison of dynamic change of follicle development among buffalo heifers synchronized by PmPGMH in low (0.3 mg/kg), medium (0.4 mg/kg) and high (0.5 mg/kg) doses of mifepristone, we found that the follicle diameter at maximal size in dose of 0.3 mg/kg was greatest among those in other doses. A comprehensive analysis on estrous rate, ovulation rate, conception rate and dynamic of follicle development showed that the dose of 0.3 mg/kg of mifepristone was the best in PmPGMH method for synchronization and TAI (Fig. 3).

Optimization of Insemination time in PmPGMH Method

Table 3 results showed that, after 24 h mifepristone injection artificial insemination time estrus rate (88.89%), ovulation rate (83.33%) and conception rate (50.00%) were higher (P < 0.05) than all other groups (Table 3), However follicle growth rate and Ovulation largest follicle diameter (Fig. 4) were not significantly different in all five groups. Considering the economic and management, we suggested that the best AI time in buffalo heifers is 24 h after mifepristone injection.

Discussion

Many strategies have been performed to improve estrus and ovulation synchronization and TAI programs for buffalo. However, it failed to attain the same success in heifers (Dahlen *et al.*, 2003; Lamb *et al.*, 2006). The

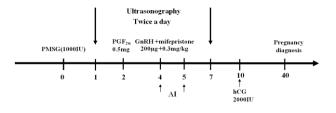


Fig. 1: Schematic diagram of PmPGMH method

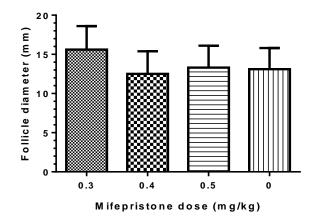


Fig. 2: Follicle diameters (mean \pm SEM) at maximal size of buffalo heifers synchronized by PmPGMH in different doses of mifepristone

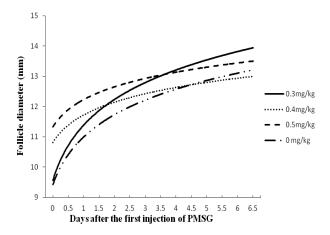


Fig. 3: Dynamic change (diameters, in mean \pm SEM) of follicle development in buffaloes synchronized by PmPGMH in low (0.3 mg/kg), medium (0.4 mg/kg) and high (0.5 mg/kg) doses of mifepristone during the first treatment to the day that a follicle grown in maximal diameter. Values in both X- and Y-coordinates were transformed in logarithm

present study developed a new synchronization protocol combined with mifepristone based on the conventional PmPG protocol named PmPGMH–TAI. The effects of different mifepristone doses, as well as the AI time on different reproductive events were also described in Chinese crossbred buffalo heifers in this new protocol.

Table 1: Onset of standing heat, Estrus duration, onset of ovulation and follicle diameter of buffalo heifers synchronized by PmPGMH method in doses of 0, 0.3, 0.4 and 0.5 (mg/kg) of mifepristone

Mifepristone dose	Onset of standing estrus	Estrus duration after onset of	Time of ovulation after second GnRH (h)	Maximum follicle
(mg/kg)	after second GnRH (h)	standing estrus (h)		diameter (mm)
0.3 (n=19)	8.3 ± 0.9^{a}	15.2 ± 0.4^{a}	24.2 ± 1.2^{a}	11.4 ± 1.4^{a}
0.4 (n=19)	8.1 ± 0.5^{a}	15.3 ± 0.5^{a}	23.5 ± 1.5^{a}	10.7 ± 1.8^{a}
0.5 (n=20)	9.0 ± 0.7^{a}	15.4 ± 0.8^{a}	24.3 ± 0.8^{a}	11.4 ± 1.5^{a}
0 (n=18)	8.0 ± 0.6^a	16.2 ± 0.6^a	24.2 ± 1.3^{a}	11.1 ± 1.6^{a}

Note: The superscripts in different small letters in the same column indicate statistical difference P < 0.05

Table 2: Estrus, ovulation and conception of buffalo heifers synchronized by PmPGMH method in doses of 0, 0.3, 0.4 and 0.5 (mg/kg) of mifepristone

Mifepristone dose	No of animals	No of animals in	No of animals in heat (%)	No of animals	No of animals pregnant (%)	No of animals with
(mg/kg)	treated	silent estrus (%)		ovulated (%)		follicle cysts(%)
0.3	19	1 (5.3 ^b)	16 (84.2 ^a)	17 (89.5°)	9 (47.4 ^b)	1 (5.26 ^a)
0.4	19	4 (21.1 ^{ab})	14 (73.7 ^{ab})	13 (68.4 ^{ab})	6 (31.6 ^{ab})	$2(10.5^{a})$
0.5	20	$6(30.0^{a})$	11 (55.0 ^b)	11 (55.0 ^b)	4 (20.0 ^a)	$2(10.0^{a})$
0	18	$3(16.6^{ab})$	12 (66.7 ^{ab})	11 (72.2 ^b)	6 (33.3 ^{ab})	2(11.1 ^a)

Note: The superscripts in different small letters in the same column indicate statistical difference P < 0.05

Table 3: Estrus, ovulation and conception and follicle dynamics of buffalo heiferssynchronized by PmPGMH method in different AI time after mifepristone injection

AI time after mifepristone (h) (n=79)	No of animals treated	No of animals in heat (%)	No of animals ovulated (%)	No of animals pregnant (%)
16	15	9 (60.00 ^{ab})	8 (53.33 ^{ab})	3 (20.00 ^{ab})
20	14	11 (78.57 ^{ab})	$10(71.42^{ab})$	5 (35.71 ^{ab})
24	18	16(88.89 ^a)	15 (83.33 ^a)	9 (50.00°)
28	17	11 (64.71 ^{ab})	11 (64.71 ^{ab})	4 (23.53 ^{ab})
32	15	7 (46.67 ^b)	6 (40.00 ^b)	2 (13.33 ^b)

Note: The superscripts in different small letters in the same column indicate statistical difference P < 0.05

Previous findings suggested that PMSG combination with Crestar implants resulted in higher estrus induction of more than 80% and conception rates of 45–66.67% in true anestrous cattle and buffaloes (Nayak *et al.*, 2009; Kumar *et al.*, 2010; Jerome *et al.*, 2016). However, other findings recorded that pregnancy rate was reduced with the increase in dosage of PMSG in dairy cows, it was significantly lower (25%) in the 4 IU/kg dose group than that of 2 and 2.5 IU/kg dose groups (71.4 and 66.7%, respectively). Moreover, higher dose of PMSG resulted in failure of ovulation due to excessive follicular development (Fu *et al.*, 2013).

High progesterone level in follicular fluid resulted in preventing the onset of LH surge, a primary hormone responsible for ovulation, by negative feedback upon the hypothalamus or pituitary thus leading to formation of follicular cysts with the incidence of 2.8% in buffalo heifers to 4.2% in buffalo cows (Luktuke and Arora, 1972; Khan et al., 2011). This follicles abnormally produced high levels of estradiol, which may interfere with embryo development (Barrett et al., 2004; Kiewisz et al., 2011). Mifepristone, as antiprogestogen, can block the effect of progesterone and induce ovulation (Parra et al., 2000; Carvalho et al., 2014; Diantonio et al., 2015; Check et al., 2016). Moreover, A desirable effect on ovulation synchronization was reported after GnRH Injection at 20 and 24 h after progesterone removal, with LH surge created 2 h after its injection in season or out of the reproductive

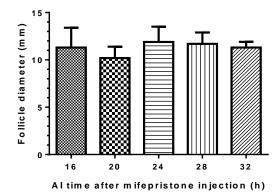


Fig. 4: Follicle diameters (mean \pm SEM) at maximal size of buffalo heifers synchronized by PmPGMH in different AI times after mifepristone injection

season (Reyna *et al.*, 2010) The results in present study showed that buffaloes treated with Mifepristone (0.3 mg/kg body weight, at the same time with GnRH injection) combined with PmPG method reduced follicular cyst rate (5.0%, 1/19), compared with PmPG treatment that induced relatively higher follicular cyst rate (11.1%, 2/18), thus improved estrous rate (84.2%, 16/19), ovulation rate (89.5%, 17/19) and pregnancy rate (47.4%, 9/19) in buffalo heifers

Silent estrus represented the main obstacle threat in productivity and reproductive efficiency in water

buffaloes because of prolonged calving intervals and great loss in milk production (Awasthi *et al.*, 1998; Madam, 1998; Prakash, 2002). It was reported that 30% of buffaloes ovulated without obvious estrous signs after PGF2 treatment (Awasthi *et al.*, 2007). PGF_{2 α} treatment in previous studies showed reduction in the concentration of progesterone in plasma in cyclic Murrah–cross buffaloes (Brito *et al.*, 2002). So declined silent estrous rate in present study can be explained due to the antiprogestogenic mechanism of mifepristone to reduce p4 concentration in plasma after mifepristone treatment. The present study showed that buffaloes treated with mifepristone (0.3 mg/kg body weight) combined with PmPG method decreased silent estrus rate (5.3%) in comparison with PmPG treatment (16.6%).

FTAI have been used to improve pregnancy rates by delaying AI of anestrous females 20 to 24 h after the predicted time (Thomas *et al.*, 2014). Previous studies in cows synchronized with PMSG in combination with PGF2α and artificially inseminated 12 and 20 h after standing heat resulted in pregnancy rate of 66.7% (Fu *et al.*, 2013). Present study revealed that PmPGMH synchronization method with TAI at 24 h after mifepristone injection resulted in the best estrus rate (88.89%), ovulation rate (83.33%) and conception rate (50%) in buffalo heifers.

Considering all the reproduction performance, it can be concluded that PmPGMH protocols showed the best synchronization achievements of stage of the estrous cycle 84.2%, induced greater ovulation rate 89.5% and improved pregnancy rate (47.4%) compared with PmPG method (33.3%) after TAI in swamp and river crossbred buffalo heifers undergoing commercial milking.

Conclusion

A new synchronization and fixed-time artificial insemination method named PmPGMH have been developed in crossbred buffalo heifers (Fig. 1). Heifers synchronized with PmPGMH in mifepristone dose of 0.3 mg/kg resulted in better synchronization than other methods. Moreover, AI at 24 h of mifepristone injection is the best artificial insemination time for heifers. The PmPGMH estrus synchronization method is as the following: (1) Crossbred buffalo heifers were subjected to intramuscular injection (I/M) of 100OIU PMSG on day 0, followed by an injection of PGF2 α (0.5 mg) on day 2; (2) Simultaneous injection of second GnRH (400 µg) and one injection of mifepristone (0.4 mg/kg) on day 4; (3) Artificial insemination was performed in the afternoon of day 5, which is 24 h after mifepristone injection; (4) I/M injection of hCG (2000 IU) in the afternoon of day 10; (5) pregnancy diagnoses was performed on day 40 after AI.

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Reference

- Akhtar, M.S., S. Ullah, L.A. Lodhi, Z.I. Qureshi and I. Ahmad, 2014. Effect of treatment with or without estradiol after Ovsynch protocols at timed AI on the pregnancy rate in lactating buffaloes. *Buff. Bull.*, 33: 184–191
- Arshad, U., A. Qayyum, M. Hassan, A. Husnain, A. Sattar and N. Ahmad, 2017. Effect of resynchronization with GnRH or progesterone (P4) intravaginal device (CIDR) on Day 23 after timed artificial insemination on cumulative pregnancy and embryonic losses in CIDR-GnRH synchronized Nili-Ravi buffaloes. *Theriogenology*, 103: 104-109
- Awasthi, M.K., F.S. Kavani, G.M. Siddiquee, N.P. Sarvaiya and H.J. Derashri, 2007. Is slow follicular growth the cause of silent estrus in water buffaloes? *Anim. Reprod. Sci.*, 99: 258–268
- Awasthi, M.K., R.P. Tiwari and G.R. Pangaonkar, 1998. Induction of estrus and fertility with low dose of prostaglandin F2 alpha in subestrus buffaloes. *Ind. J. Anim. Sci.*, 68: 1049–1050
- Barrett, D.M.W., P.M. Bartlewski, M. Batista–Arteaga, A. Symington and N.C. Rawlings, 2004. Ultrasound and endocrine evaluation of the ovarian response to a single dose of 500 IU of eCG following a 12–day treatment with progestogen–releasing intravaginal sponges in the breeding and nonbreeding seasons in ewes. *Theriogenology*, 61: 311–327
- Baruselli, P.S., 2001. Control of Follicular Development Applied to Reproduction Biotechnologies in Buffalo, pp. 128–146. Anais, Eboli
- Baruselli, P.S., 1994. Basic requirements for artificial insemination and embryo transfer in buffaloes. *Buff. J.*, 2: 53–60
- Baruselli, P.S., V.H. Barnabe, R.C. Barnabe, J.A. Visintin, J.R. Molero–Filho and R. Porto, 2001. Effect of Body Condition Score at Calving on Postpartum Reproductive Performance in Buffalo. Buff. J., 17: 53–65
- Berber, R.C.D.A., E.H. Madureira and P.S. Baruseli, 2002. Comparison of two Ovsynch protocols (GnRH versus LH) for fixed timed insemination in buffalo (*Bubalus bubalis*). *Theriogenology*, 57: 1421–1430
- Brito, L.F.C., R. Satrapa, E.P. Marson and J.P. Kastelic, 2002. Efficacy of PGF 2α to synchronize estrus in water buffalo cows (*Bubalus bubalis*) is dependent upon plasma progesterone concentration, corpus luteum size and ovarian follicular status before treatment. *Anim. Reprod. Sci.*, 73: 23–35
- Caesar, N.K., S.N. Shukla, O.P. Shrivastava, S. Agrawal and R.G. Agrawal, 2011. Studies on fertility response in anoestrus buffaloes using a modified CIDR-based synchronization protocol. *Buff. Bull.*, 30: 184–187
- Carvalho, N.A.T., J.G. Soares, D.C. Souza, J.R. Maio, J.N. Sales, J.B. Martins, R.C. Macari, M.J. D'Occhio and P.S. Baruselli, 2017. Ovulation synchronization with estradiol benzoate or GnRH in a timed artificial insemination protocol in buffalo cows and heifers during the nonbreeding season. *Theriogenology*, 87: 333–338
- Carvalho, N.A.T., J.G. Soares and P.S. Baruselli, 2016. Strategies to overcome seasonal anestrus in water buffalo. *Theriogenology*, 86: 200–206
- Carvalho, N.A.T., J.G. Soares, D.C. Souza, F.S. Vannucci, R. Amaral, J.R. Maio, J.N. Sales, M.F.S. Filho and P.S. Baruselli, 2014. Different circulating progesterone concentrations during synchronization of ovulation protocol did not affect ovarian follicular and pregnancy responses in seasonal anestrous buffalo cows. *Theriogenology*, 81: 490–495
- Carvalho, N.A.T., J.G. Soares, R.M. Filho, Porto, L.U. Gimenes, D.C. Souza, M. Nichi, J.S. Sales and P.S. Baruselli, 2013. Equine chorionic gonadotropin improves the efficacy of a timed artificial insemination protocol in buffalo during the nonbreeding season. Theriogenology, 79: 423–428

- Check, J.H., G. Diantonio, A. Diantonio and M. Duroseau, 2016. The progesterone receptor antagonist mifepristone does not lower serum progesterone induced blocking factor (PIBF) in the presence of progesterone. Clin. Exp. Obstet. Gynecol., 43: 189–191
- Dahlen, C.R., G.C. Lamb, C.M. Zehnder, L.R. Miller and A. Dicostanzo, 2003. Fixed-time insemination in peripuberal, lightweight replacement beef heifers after estrus synchronization with PGF2alpha and GnRH. *Theriogenology*, 59: 1827–1837
- Diantonio, G., J.H. Check, A. Diantonio and M. Duroseau, 2015. The Progesterone (P) Receptor Antagonist Mifepristone Does Not Lower Serm Progeserone Induced Blocking Factor (PIBF) in the Presence of P. Fertil. Steril., 103: 18–18
- Fu, S.B., H.L. Zhang, H. Riaz, S. Ahmad, X.M. Wang, X. Li, G.H. Hua, X.R. Liu, A.Z. Guo and L.G. Yang, 2013. Effects of different doses of PMSG on reproductive performance in Chinese Holstein dairy cows. Pak. Vet. J., 33: 209–215
- Garoussi, M.T., N. Farzaneh, E. Gallehdar and M. Mohri, 2012. Reproductive performance in out-of-breeding season of fatty ewes using implant norgestomet with or without PMSG. *Trop. Anim. Hlth. Prod.*, 44: 965–968
- Gimenes, L.U., N.A.T. Carvalho, M.F.S. Filho, F.S. Vannucci, J.R.S. Torres–Júnior, H. Ayres, R.M. Ferreira, L.A. Trinca, E.S. Sartorelli, C.M. Barros, M.P. Beltran, G.P. Nogueira, R.J. Mapletoft and P.S. Baruselli, 2011. Ultrasonographic and endocrine aspects of follicle deviation, and acquisition of ovulatory capacity in buffalo (*Bubalus bubalis*) heifers. *Anim. Rep. Sci.*, 123: 175–179
- Giuseppe, C., N. Gianluca, G. Bianca, G. Giorgio, P. Alberto, D.P. Rossella, M.J. D'Occhio and Z. Luigi, 2005. Embryonic mortality in buffaloes synchronized and mated by AI during the seasonal decline in reproductive function. *Theriogenology*, 63: 2334–2340
- Greyling, J.P.C. and C.H.V. Niekerk, 1991. Different synchronization techniques in Boer goat does outside the normal breeding season. Small Rumin. Res., 5: 233–243
- Haider, M.S., M. Hassan, A.S. Khan, A. Husnain, M. Bilal, J.R. Pursley and N. Ahmad, 2015. Effect of timing of insemination after CIDR removal with or without GnRH on pregnancy rates in Nili–Ravi buffalo. Anim. Reprod. Sci., 163: 24–29
- Jainudeen, M.R. and E.S.E. Hafez, 2016. Cattle and Buffalo. In: Reproduction in Farm Animals, 7th edition, pp: 157–171. John Wiley and sons, Inc.
- Jerome, A., S.K. Srivastava and R.K. Sharma, 2016. Study on follicular characteristics, hormonal and biochemical profile in norgestomet+PMSG treated acyclic buffaloes. *Iran. J. Vet. Res.*, 17: 247–252
- Khan, F.A., G.K. Das, M. Pande, M.K. Pathak and M. Sarkar, 2011. Biochemical and hormonal composition of follicular cysts in water buffalo (Bubalus bubalis). Anim. Reprod. Sci., 124: 61–64
- Kiewisz, J., M.M. Kaczmarek, E. Morawska, A. Blitek, W. Kapelanski and A.J. Ziecik, 2011. Estrus synchronization affects WNT signaling in the porcine reproductive tract and embryos. *Theriogenology*, 76: 1684–1694
- Kumar, H., N. Bhooshan, M.K. Patra and M.C. Yadav, 2010. Treatment with progestagen and PMSG to prevent prolonged anestrus in buffaloes. *Ind. J. Anim. Sci.*, 80: 623–625
- Lamb, G.C., J.E. Larson, T.W. Geary, J.S. Stevenson, S.K. Johnson, M.L. Day, R.P. Ansotegui, D.J. Kesler, J.M. Dejarnette and D.G. Landblom, 2006. Synchronization of estrus and artificial insemination in replacement beef heifers using gonadotropin–releasing hormone, prostaglandin F2 alpha, and progesterone. J. Anim. Sci., 84: 3000–3009
- Liu, Q., H. Li, Z.U. Rehman, X. Dan, X. Liu, D. Bhattarai and L. Yang, 2016. The efficacy of an inhibin DNA vaccine delivered by attenuated Salmonella choleraesuis on follicular development and ovulation responses in crossbred buffaloes. Anim. Reprod. Sci., 172: 76–82
- Lokeshwar, R.R., 1990. Efficiency of semen production in buffalo bulls. Recent Adv. Buff. Res. 3: 87–88
- Lu, Y., Y. Liao, M. Zhang, B. Yang, X. Liang, X. Yang, S. Lu, Z. Wu, H. Xu and Y. Liang, 2015. A field study on artificial insemination of swamp and crossbred buffaloes with sexed semen from river buffaloes. *Theriogenology*, 84: 862–867
- Luktuke, S.N. and R.L. Arora, 1972. Studies on cystic degeneration of ovaries in buffalo females. *Ind. Vet. J.*, 49: 146–150

- Madam, M.L., 1998. Status of Reproduction in Female Buffalo, pp: 89–100.
 Buffalo Production and Health: A Compendium of Last EST Research Information based on Indian Studies. ICAR Publication, New Delhi, India
- Mehaisen, G.M.K., J.S. Vicente, R. Lavara and M.P. Viudes–De–Castro, 2005. Effect of eCG dose and ovulation induction treatments on embryo recovery and in vitro development post–vitrification in two selected lines of rabbit does. Anim. Reprod. Sci., 90: 175–184
- Monteiro, B.M., D.C. Souza, G. Vasconcellos, N.A.T. Carvalho and P.S. Baruselli, 2018. Effect of season on dairy buffalo reproductive performance when using P4/E2/eCG-based fixed-time artificial insemination management. *Theriogenology*, 119: 275–281
- Muth–Spurlock, A.M., D.H. Poole and C.S. Whisnant, 2016. Comparison of pregnancy rates in beef cattle after a fixed–time AI with once– or twice–used controlled internal drug release devices. *Theriogenology*, 85: 447–451
- Nayak, V., R.G. Agrawal, O.P. Srivastav and M.S. Thakur, 2009. Induction of estrus in true anestrus buffaloes using Crestar implants alone and in combination with PMSG. Buff. Bull., 28: 51–54
- Parra, J., B. Cantabrana and A. Hidalgo, 2000. Mechanism of mifepristone–induced spasmolytic effect on isolated rat uterus. *Life Sci.*, 66: 2563–2569
- Prakash, B.S., 2002. *Influence of Environment on Animal Reproduction*, pp: 34–47. Invited Paper: National Workshop on Animal Climate Interaction, held at Izatnagar, India
- Presicce, G.A., E.M. Senatore, G.D. Santis and A. Bella, 2005. Follicle turnover and pregnancy rates following oestrus synchronization protocols in Mediterranean Italian buffaloes (*Bubalus bubalis*). *Reprod. Domest. Anim.*, 40: 443–447
- Rajesh, G., A. Paul, S.R. Mishra, J. Bharati, N. Thakur, T. Mondal, S. Soren, S. Harikumar, K. Narayanan, V.S. Chouhan, S. Bag, B.C. Das, G. Singh, V.P. Maurya, G.T. Sharma and M. Sarkar, 2017. Expression and functional role of bone morphogenetic proteins (BMPs) in cyclical corpus luteum in buffalo (*Bubalus bubalis*). Gen. Comp. Endocrinol., 240: 198–213
- Rensis, F.D. and F. López–Gatius, 2007. Protocols for synchronizing estrus and ovulation in buffalo (*Bubalus bubalis*): a review. *Theriogenology*, 67: 209–216
- Reyna, J., P.C. Thomson, G. Evans and W.M. Maxwell, 2010. Synchrony of ovulation and follicular dynamics in merino ewes treated with GnRH in the breeding and non-breeding seasons. *Reprod. Domest. Anim.*, 42: 410–417
- Rostami, B., A. Niasari–Naslaji, M. Vojgani, D. Nikjou, H. Amanlou and A. Gerami, 2011. Effect of eCG on early resumption of ovarian activity in postpartum dairy cows. *Anim. Reprod. Sci.*, 128: 100–106
- Singh, J., A.S. Nanda and G.P. Adams, 2000. The reproductive pattern and efficiency of female buffaloes. *Anim. Reprod. Sci.*, 60: 593–604
- Souza, A.H., S. Viechnieski, F.A. Lima, F.F. Silva, R. Araújo, G.A. Bó, M.C. Wiltbank and P.S. Baruselli, 2009. Effects of equine chorionic gonadotropin and type of ovulatory stimulus in a timed–AI protocol on reproductive responses in dairy cows. *Theriogenology*, 72: 10–21
- Thomas, J.M., S.E. Poock, M.R. Ellersieck, M.F. Smith and D.J. Patterson, 2014. Delayed insemination of non–estrous heifers and cows when using conventional semen in timed artificial insemination. *J. Anim. Sci.*, 92: 4189–4197
- Turk, G., S. Gur, M. Sonmez, T. Bozkurt, E.H. Aksu and H. Aksoy, 2008. Effect of exogenous GnRH at the time of artificial insemination on reproductive performance of Awassi ewes synchronized with progestagen–PMSG–PGF2alpha combination. *Reprod. Domest. Anim.*, 43: 308–313
- Ustuner, B., U. Gunay, Z. Nur and H. Ustuner, 2007. Effects of long and short-term progestagen treatments combined with pmsg on oestrus synchronization and fertility in awassi ewes during the breeding season. *Acta Vet. Brno*, 76: 391–397
- Yindee, M., M. Techakumphu, C. Lohachit, S. Sirivaidyapong, A. Na–Chiangmai, H. Rodriguez–Martinez, G.C.V.D. Weyden and B. Colenbrander, 2011. Follicular dynamics and oestrous detection in thai postpartum swamp buffaloes (*Bubalus bubalis*). Reprod. Domest. Anim., 46: 91–96

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